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# Statistical SERS spectroscopy based on concentration-encoded intensity distributions for deep-learning-assisted quantitation

Meng Zhang<sup>1†</sup>, Hui Ding<sup>2†</sup>, Luyuan Xie<sup>3</sup>, Chaowei Sun<sup>4</sup>, Shiyu Zhang<sup>2</sup>, Yue Wei<sup>1</sup>, Di Zheng<sup>1</sup>, Ferruccio Pisanello<sup>5</sup>, Longkun Yang<sup>1\*</sup>, Hongxing Xu<sup>4\*</sup> and Zhipeng Li<sup>1\*</sup>

**Abstract:** Reliable quantitative SERS analysis is limited by strong intensity fluctuations arising from stochastic molecule-substrate-light interactions. Here, we reveal that SERS intensity distributions carry concentration-dependent statistical information, exhibiting a characteristic evolution from an asymmetric long-tailed profile at low concentration to a nearly symmetric Gaussian-like pattern at high concentration. This concentration-dependent evolution originates from stochastic molecular adsorption at plasmonic “hot spots” and can be reproduced by Poisson-statistics-based Monte Carlo simulations. On this basis, we propose a statistical SERS strategy for concentration-level quantitation, in which concentration-encoded intensity distribution patterns are identified using a ResNet4 deep learning model. Accurate concentration identification of R6G is achieved, and the generality of this method is further demonstrated by the analysis of methamphetamine in methanol, thiram in bean sprout extract, and rosiglitazone maleate in serum. These results transform intrinsic SERS intensity fluctuations from a limitation of conventional intensity-based quantitation into useful statistical fingerprints, which certainly deepens our understanding about the stochastic nature of SERS and provides a new route toward reliable SERS quantitation for chemical and biological analysis.

**Keywords:** statistical spectroscopy; surface-enhanced Raman scattering; poisson distribution; quantitative detection

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## 1 Introduction

Statistical spectroscopy has emerged as an interdisciplinary field that integrates statistical methodologies with spectroscopic techniques, offering powerful solutions for analyzing complex spectral data across physics, chemistry, and astronomy<sup>1–5</sup>. A particularly groundbreaking application lies in quantitative analysis at the single-molecule level<sup>6–8</sup>, where Nilsson et al. pioneered digital counting quantitation by converting analog fluorescence signals into binary digital outputs (0/1)<sup>9</sup>. This approach leverages Poisson statistics, where quantitation accuracy scales with the square root of event counts<sup>7,9</sup>, establishing a robust framework for ultra-sensitive quantitation.

Compared with the fluorescence, Raman spectroscopy offers rich vibrational fingerprints for molecular identification, which is particularly advantageous for analyzing in complex matrices<sup>10–16</sup>. With plasmonic enhancement, surface-enhanced Raman scattering (SERS) can further improve detection sensitivity down to the single-molecule level<sup>17–19</sup>. While, SERS signals are highly sensitive to stochastic molecule-substrate-light interactions<sup>12,20–22</sup>. The random adsorption of molecules at plasmonic “hot spots” and the heterogeneous enhancement distribution of SERS substrates lead to pronounced intensity fluctuations<sup>23,24</sup>, making conventional intensity-based quantitation challenging. Recently, a breakthrough on quantitative SERS has been

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made by introducing statistical digital counting method<sup>25,26</sup>. By gauging SERS probability rather than absolute intensity, quantitation at single-molecule concentration can even be achieved with high precision. It has demonstrated practical applicability in fields such as environmental protection and food safety, which indicates the rising of statistical SERS spectroscopy<sup>27–31</sup>. Beyond binary event counting, SERS intensity distributions contain richer statistical information associated with both molecular-number fluctuations and “hot-spot” dependent enhancement variations<sup>32,33</sup>. The former is related to the number of molecules contributing to each SERS measurement<sup>34</sup>, whereas the latter reflects the enhancement characteristics of the SERS substrate<sup>35</sup>. Therefore, deciphering how SERS intensity distributions evolve with analyte concentration may provide a new paradigm for quantitative SERS analysis.

In this work, we propose a statistical SERS quantitation strategy based on deep-learning-assisted identification of concentration-encoded SERS intensity distributions (Fig. 1). We first reveal that SERS intensity distributions exhibit a clear concentration-dependent evolution. As the R6G concentration increases from  $1 \times 10^{-11}$  M to  $1 \times 10^{-7}$  M, the intensity distribution gradually evolves from an asymmetric long-tailed profile to a nearly symmetric Gaussian-like pattern. This evolution can be explained by stochastic molecular adsorption at plasmonic “hot spots” and is reproduced by Poisson-statistics-based Monte Carlo simulations. The subtle but concentration-specific distribution features can be effectively identified by the ResNet4 deep learning model, enabling accurate concentration-level quantitation of R6G with an accuracy up to 100%. The generality and practical applicability of this strategy are further demonstrated by the analysis of methamphetamine in methanol (1 ppb–10 ppm), thiram in bean sprout extract ( $1 \times 10^{-8}$  M– $1 \times 10^{-6}$  M), and rosiglitazone maleate in serum ( $1 \times 10^{-7}$  M– $1 \times 10^{-5}$  M). This statistical strategy transforms intrinsic SERS intensity fluctuations from an obstacle in conventional

intensity-based quantitation into useful concentration-encoded fingerprints, providing a new route toward reliable SERS quantitation in chemical and biological analysis.

## 2 Results and discussion

### 2.1 Statistical distribution characteristics of SERS intensity

Statistical SERS quantitation requires a large number of spectra acquired from substrates with good spatial uniformity. To meet this requirement, well-ordered silver nanoparticle arrays were fabricated using the Langmuir-Blodgett (LB) technique, as described in the Experimental section<sup>36,37</sup>. Scanning electron microscopy (SEM) images (Fig. S3 in the Supplementary information) show that the silver nanoparticles (~60 nm in diameter) are closely packed and uniformly distributed over a micrometer-scale area, providing a suitable platform for SERS mapping measurements. R6G was selected as a model analyte. The SERS samples were prepared by immersing the substrates in R6G solutions with different concentrations for 2 h, followed by drying under ambient conditions. Unless otherwise stated, SERS measurements were performed under 532 nm laser excitation with a power of 7.5  $\mu$ W and an integration time of 1 s. The mapping area was  $30 \times 30 \mu\text{m}^2$  with a step size of 1  $\mu\text{m}$ . Figure 2(a) shows the SERS mapping images of R6G at  $612 \text{ cm}^{-1}$  over the concentration range from  $1 \times 10^{-11}$  M to  $1 \times 10^{-7}$  M. At each concentration, 100 pixels were randomly selected from the mapping image, and the corresponding SERS intensities at  $612 \text{ cm}^{-1}$  are plotted in Fig. 2(b). The average SERS intensity generally increased with increasing R6G concentration, and the fitted calibration curves gave  $R^2$  values of approximately 0.90 (Fig. S4(a)). However, large intensity fluctuations were observed at each concentration. For example, the relative standard deviation (RSD) of the SERS intensity at  $1 \times 10^{-9}$  M reached 37.9%. Since

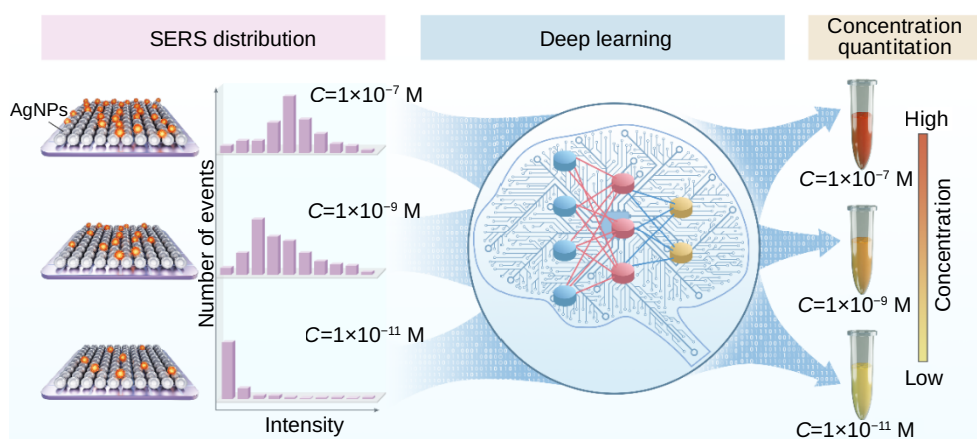


Fig. 1 | The scheme of SERS quantitation via deep learning identified statistical distributions of intensity.

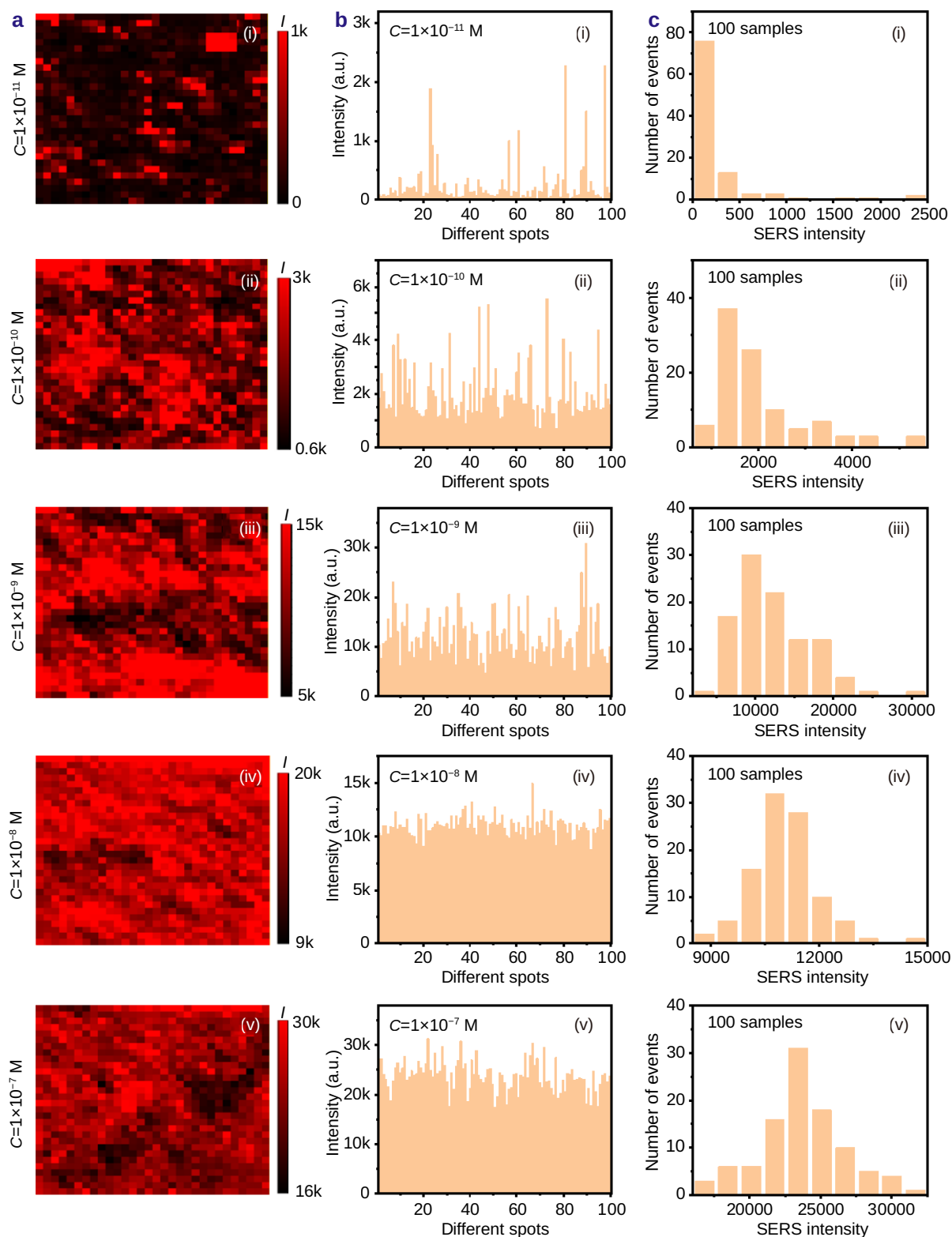


Fig. 2 | Statistical distributions of R6G SERS intensity. (a) SERS mapping images of R6G acquired at  $612 \text{ cm}^{-1}$  at different concentrations. Panels (i)–(v) correspond to  $1 \times 10^{-11} \text{ M}$ ,  $1 \times 10^{-10} \text{ M}$ ,  $1 \times 10^{-9} \text{ M}$ ,  $1 \times 10^{-8} \text{ M}$ , and  $1 \times 10^{-7} \text{ M}$ , respectively. (b) SERS intensities at  $612 \text{ cm}^{-1}$  extracted from 100 randomly selected pixels in the corresponding mapping images (a). (c) Statistical histograms of SERS intensity in (b).

conventional intensity-based quantitation uses the SERS intensity as the sole quantitative metric, such large signal fluctuations inevitably reduce the precision of concentration determination. Moreover, the calibration curves obtained from different batches exhibited different slopes and intercepts (Fig. S4(b)), further indicating that robust quantitation based solely on absolute SERS intensity remains challenging.

To overcome this limitation, we analyzed the statistical distribution of SERS intensities rather than individual intensity values. At each R6G concentration, the SERS intensity values at 612 cm<sup>-1</sup> were uniformly divided into 10 bins from  $I_{\min}$  to  $I_{\max}$ , where  $I_{\min}$  and  $I_{\max}$  were determined from the corresponding full mapping dataset. The number of SERS events falling into each bin was then counted to construct a statistical intensity histogram. The obtained intensity distributions are shown in Fig. 2(c). Interestingly, the distribution patterns exhibited a clear dependence on analyte concentration. At  $1 \times 10^{-11}$  M, most SERS events accumulated in the first bin, forming an asymmetric long-tailed, Pareto-like distribution (panel i)<sup>38,39</sup>. As the concentration increased, the population of SERS events gradually shifted toward higher-intensity bins (panels ii–iv), and finally evolved into a nearly Gaussian-like distribution at  $1 \times 10^{-7}$  M (panel v). To quantitatively describe this concentration-dependent evolution, the skewness of the raw SERS intensity data was calculated at each concentration. As shown in Table S1, the skewness decreased from 3.515 at  $1 \times 10^{-11}$  M to 0.086 at  $1 \times 10^{-7}$  M, confirming that the intensity distribution gradually evolved from a highly asymmetric right-skewed profile to a nearly symmetric one with increasing analyte concentration.

The concentration-dependent evolution of SERS intensity distributions can be understood from the stochastic combination between molecules and plasmonic “hot spot”. The number of molecules contributing to each SERS spectrum can be described by Poisson statistics. The probability of measuring ( $n$ ) molecules in one spectrum follows the relation:

$$p(n, \alpha) = \frac{\alpha^n}{n!} e^{-\alpha}, \quad (1)$$

where  $\alpha$  represents the average number of molecules per spectrum at a given analyte concentration. The electromagnetic “hot spot” was modeled using a coupled Ag nanoparticle dimer with a diameter of 60 nm and a gap size of 2 nm, as shown in Fig. S5(a). Assuming a total number of spectral samplings  $N_s$ , the number of spectra containing  $n$  molecules is given by  $N_s p(n)$ . In the Monte Carlo simulations, these  $n$  molecules were randomly distributed on the surface of the nanoparticle dimer structure. For each sampling event containing  $n$  molecules, the SERS intensity will be proportional to

$$I_j \sim \sum_{i=1}^n |E_i|^4, \quad (2)$$

where  $E_i$  is the local electric field at the location of the  $i$ th molecule. For each  $\alpha$  value, corresponding to a given analyte concentration in the experiment, a set of simulated SERS intensities  $\{I_1, I_2, \dots, I_{N_s}\}$  can be obtained by performing  $N_s$  times Monte Carlo simulations. Histograms were then constructed to analyze the simulated intensity distributions. As shown in Fig. S5(b), when  $\alpha$  increased from 40 to 2000, the simulated intensity distribution gradually evolved from a Pareto-like long-tailed pattern to a near-Gaussian profile, well reproducing the experimental observations. To further examine the influence of near-field enhancement, additional simulations were performed using Ag nanoparticle dimers with different gap sizes. As shown in Fig. S5(c), reducing the gap size from 2 nm to 1 nm generated a stronger and more localized electromagnetic field in the junction region. Accordingly, the simulated SERS intensities increased, and the intensity distributions become broader and more skewed, especially at low and intermediate  $\alpha$  values. These results indicate that the gap size or near-field enhancement strength can affect the absolute intensity scale, distribution width and detailed distribution profile. Nevertheless, the overall  $\alpha$ -dependent evolution remains unchanged: the intensity distribution evolves from a skewed long-tailed pattern at low  $\alpha$  to a more symmetric profile at high  $\alpha$ . Therefore, the near-field enhancement modulates the detailed intensity distributions, but does not alter the fundamental statistical mechanism responsible for the concentration-dependent evolution of SERS intensity distributions.

## 2.2 SERS intensity distributions identified by deep learning model

To establish statistical SERS intensity distributions as a reliable quantitation metric, their reproducibility should first be verified. We calculated the Pearson correlation coefficients between SERS intensity distributions obtained from independent measurements at the same concentration. As shown in Fig. S6, strong positive correlations ( $\rho \approx 1$ ) were observed across all tested concentrations, indicating the good reproducibility of the concentration-encoded intensity distributions. This result suggests that statistical intensity distributions can serve as robust fingerprints for SERS quantitation. However, direct concentration identification based solely on Pearson correlation coefficients between an unknown distribution and standard distributions at different concentrations was found to be unreliable, especially for adjacent concentrations with highly similar distribution profiles (Fig. S7). This is because Pearson correlation mainly evaluates global linear similarity, whereas the concentration information in SERS intensity distributions is encoded in more subtle and nonlinear features, including the relative population of different intensity bins, the peak position, the distribution width, and the local variation among neighboring bins. These features are difficult to fully capture using simple

intensity calibration or correlation-based comparison.

To better extract concentration-specific distribution features, we employed the ResNet4 deep learning model to identify the statistical SERS intensity distributions. The one-dimensional convolutional layers in ResNet4 can efficiently extract local bin-level correlations among neighboring histogram bins, while the stacked residual blocks enable hierarchical feature learning from local intensity-bin variations to higher-level distribution representations. In addition, the residual connections help preserve feature information and improve optimization stability during training. Therefore, ResNet4 was selected as an appropriate classifier for distinguishing one-dimensional concentration-encoded SERS intensity distribution patterns derived from SERS mapping data<sup>40–45</sup>. The architecture of the ResNet4 model is illustrated in Fig. 3. Briefly, the input histogram data are first downsampled by a one-dimensional convolution and a max-pooling layer, then passed through four residual blocks, each with one-dimensional convolutions, batch normalization, ReLU activation, and shortcut connections. After feature extraction, adaptive average pooling generates a fixed-size feature representation, which is then fed into a fully connected layer to output the concentration class. More details of the ResNet4 model are provided in the *Experimental* section.

A sufficient number of distribution datasets is essential

for training a stable deep learning model<sup>46,47</sup>. As described in the Experimental section, we developed a random-sampling extraction strategy to generate multiple statistical distribution patterns from the SERS mapping data. For each R6G concentration, 200 distribution datasets were generated, with each histogram constructed from 100 randomly selected spectra using 10 bins. These datasets were divided into training and testing sets at a 1:1 ratio, yielding 100 training datasets and 100 testing datasets for each concentration. To evaluate the robustness of the quantitation strategy, the training and testing datasets were sourced from independent batches of SERS mapping measurements. The ResNet4 was trained using the AdamW optimizer with an initial learning rate of 0.01 and a batch size of 64. Cross-entropy loss with label smoothing was used as the loss function. The training set was further divided into training and validation subsets for model selection and early stopping, while the independent testing set was used only for final performance evaluation. As shown in Table 1, the ResNet4 model achieved 100% recognition accuracy for all five R6G concentrations, demonstrating its strong capability for concentration-level identification. For comparison, classical machine-learning methods, including KNN and SVM, were also used to classify the statistical distributions. As summarized in Table 1, ResNet4 achieved the best overall performance among the tested models. This advantage may arise from its

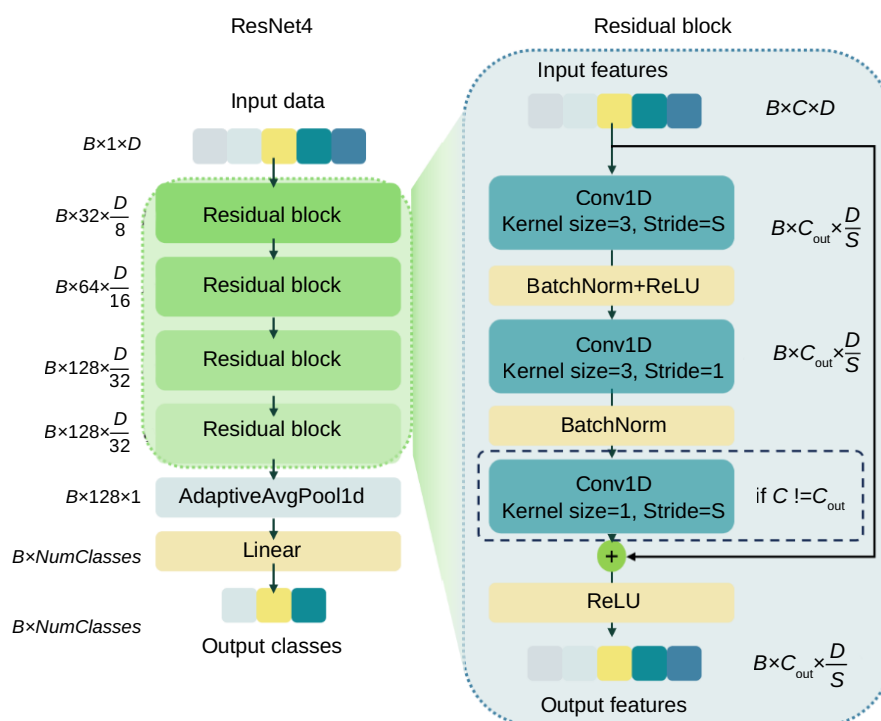


Fig. 3 | The architecture of the ResNet4 model. Left panel is the overall architecture of ResNet4, illustrating how the input data passes through multiple residual blocks, an adaptive average pooling layer, and a linear layer to ultimately output class predictions. On the right, the internal structure of a residual block is detailed, including one-dimensional convolutional layers, batch normalization layers, activation functions, and the implementation of residual connections.

ability to learn both local bin-level correlations and global distribution features from histogram inputs, whereas conventional methods based on predefined distance metrics or kernel functions are less effective in capturing subtle nonlinear differences among concentration-encoded SERS intensity distributions.

To further evaluate the concentration resolution of the proposed statistical quantitation method, we expanded the classification system by introducing four intermediate concentrations, namely  $0.5 \times 10^{-10}$  M,  $0.5 \times 10^{-9}$  M,  $0.5 \times 10^{-8}$  M, and  $0.5 \times 10^{-7}$  M, in addition to the original five concentration levels. Consistent with Table 1, the statistical intensity histograms at each concentration were constructed from 100 spectral samples using 10 bins. Each concentration contained 100 training datasets and 100 testing datasets, giving a total of 900 training datasets and 900 testing datasets. As shown in Table 2, when nine closely spaced concentration levels were included, the average recognition accuracy decreased to 77%. This decrease is expected because the statistical distribution differences between adjacent concentrations become more subtle under the current sampling density. These results indicate that finer concentration resolution requires more spectral samplings to better characterize the distribution features.

To obtain more detailed statistical information, we further increased both the number of spectral samplings and the number of histogram bins. At each concentration, 200 intensity values were randomly selected from the SERS mapping images shown in Fig. 2(a) and Fig. S8(a). These intensity values were then divided into 20 bins to generate

statistical distribution patterns, as shown in Fig. 4. It was noted that increasing the bin number did not change the fundamental concentration-dependent evolution of the distributions, but the larger number of SERS events provided richer statistical features and improved the distinguishability among adjacent concentrations. As summarized in Table 2, using 200 samples and 20 bins, the ResNet4 model achieved an average concentration identification accuracy of 100%. Further increasing the sampling number and bin number to 400 samples and 40 bins yielded a comparable average accuracy of 99% (Fig. S10). These results demonstrate that statistical distributions generated from sufficient SERS samplings can significantly improve concentration-level recognition performance. The high accuracy and robustness of this strategy highlight the potential of ResNet4-identified statistical intensity distributions for reliable SERS quantitation.

### 2.3 Applications of AI-driven statistical SERS quantitation methodology

To evaluate the applicability and generality of the proposed statistical SERS quantitation strategy, we first applied it to the analysis of methamphetamine (MA) in methanol as a proof-of-concept demonstration. MA is a representative amphetamine-type stimulant, and its rapid and sensitive detection is of practical importance<sup>48,49</sup>. The SERS spectrum, mapping images, and corresponding statistical intensity distributions of MA over the concentration range from 1 ppb to 10 ppm are shown in Fig. S11. The characteristic

Table 1 | Accuracy of concentrations identification by deep learning model (ResNet4) and machine learning models (KNN and SVM). Input data contains 5 concentrations,  $1 \times 10^{-11}$  M,  $1 \times 10^{-10}$  M,  $1 \times 10^{-9}$  M,  $1 \times 10^{-8}$  M,  $1 \times 10^{-7}$  M. At each concentration, statistical histograms are obtained from 100 spectral samples with bins number 10.

|                  | Models        | $1 \times 10^{-11}$ M | $1 \times 10^{-10}$ M | $1 \times 10^{-9}$ M | $1 \times 10^{-8}$ M | $1 \times 10^{-7}$ M | Average accuracy |
|------------------|---------------|-----------------------|-----------------------|----------------------|----------------------|----------------------|------------------|
| Deep learning    | ResNet4       | <b>100%</b>           | <b>100%</b>           | <b>100%</b>          | <b>100%</b>          | <b>100%</b>          | <b>100%</b>      |
|                  | KNN ( $K=1$ ) | 92%                   | 91%                   | 90%                  | 91%                  | 91%                  | 91%              |
| Machine learning | KNN ( $K=3$ ) | 91%                   | 90%                   | 89%                  | 90%                  | 90%                  | 90%              |
|                  | KNN ( $K=5$ ) | 88%                   | 88%                   | 87%                  | 88%                  | 89%                  | 88%              |
|                  | SVM (linear)  | 98%                   | 97%                   | 97%                  | 99%                  | 99%                  | 98%              |
|                  | SVM (rbf)     | 44%                   | 42%                   | 43%                  | 46%                  | 45%                  | 44%              |

Table 2 | Accuracy of concentrations identification by ResNet4 model, where the input data contains 9 concentration levels,  $1 \times 10^{-11}$  M,  $0.5 \times 10^{-10}$  M,  $1 \times 10^{-10}$  M,  $0.5 \times 10^{-9}$  M,  $1 \times 10^{-9}$  M,  $0.5 \times 10^{-8}$  M,  $1 \times 10^{-8}$  M,  $0.5 \times 10^{-7}$  M,  $1 \times 10^{-7}$  M. At each concentration, statistical histograms are obtained from 100/200/400 spectral samples with bins number 10/20/40.

|         | Bins number           | $1 \times 10^{-11}$ M | $0.5 \times 10^{-10}$ M | $1 \times 10^{-10}$ M | $0.5 \times 10^{-9}$ M | $1 \times 10^{-9}$ M | $0.5 \times 10^{-8}$ M | $1 \times 10^{-8}$ M | $0.5 \times 10^{-7}$ M | $1 \times 10^{-7}$ M | Average accuracy |
|---------|-----------------------|-----------------------|-------------------------|-----------------------|------------------------|----------------------|------------------------|----------------------|------------------------|----------------------|------------------|
| ResNet4 | 10 bins (100 samples) | 74%                   | 64%                     | 78%                   | 68%                    | 75%                  | 92%                    | 96%                  | 44%                    | 100%                 | 77%              |
|         | 20 bins (200 samples) | 100%                  | 100%                    | 100%                  | 100%                   | 100%                 | 100%                   | 100%                 | 100%                   | 100%                 | 100%             |
|         | 40 bins (400 samples) | 95%                   | 100%                    | 100%                  | 100%                   | 100%                 | 100%                   | 100%                 | 100%                   | 100%                 | 99%              |

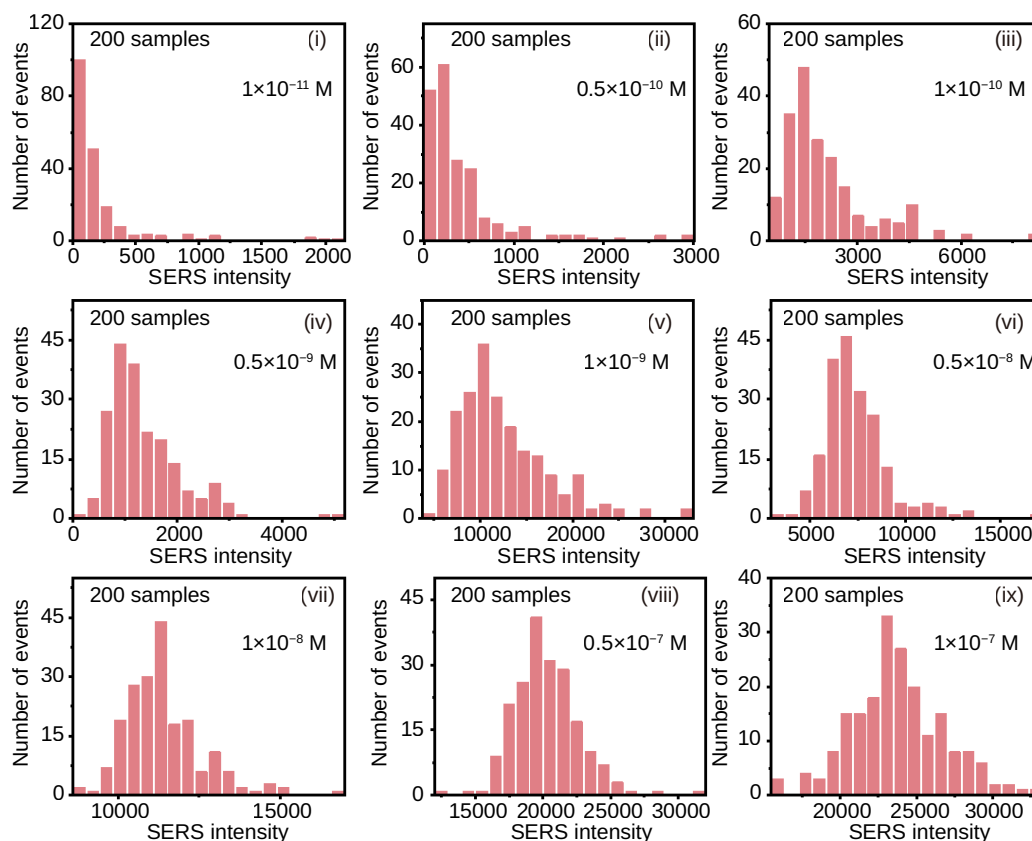


Fig. 4 | Statistical distributions of 200 SERS intensity samplings within 20 bins, where the concentrations are panel (i)  $1 \times 10^{-11}$  M, panel (ii)  $0.5 \times 10^{-10}$  M, panel (iii)  $1 \times 10^{-10}$  M, panel (iv)  $0.5 \times 10^{-9}$  M, panel (v)  $1 \times 10^{-9}$  M, panel (vi)  $0.5 \times 10^{-8}$  M, panel (vii)  $1 \times 10^{-8}$  M, panel (viii)  $0.5 \times 10^{-7}$  M, panel (ix)  $1 \times 10^{-7}$  M, respectively.

peak at  $1002 \text{ cm}^{-1}$ , assigned to the phenyl ring breathing vibration of MA, was selected to construct the intensity distributions. Using the concentration-encoded intensity distributions as inputs, the trained ResNet4 model achieved recognition accuracies higher than 98% for all tested concentrations, with an average accuracy of 99.6%, as summarized in Fig. 5(a). These results demonstrate that the proposed statistical SERS strategy is effective for concentration-level identification of small molecules in a simple solvent system.

We next evaluated the method in a more complex food matrix by detecting thiram in bean sprout extract<sup>50,51</sup>. Fresh bean sprouts were physically crushed, filtered, and centrifuged to obtain a clear extract, which was then spiked with different concentrations of thiram to simulate pesticide residues. The SERS spectrum, mapping images, and corresponding statistical intensity distributions are provided in Fig. S12. The bean sprout extract showed no obvious SERS interference at the characteristic thiram bands. The peak at  $1387 \text{ cm}^{-1}$ , assigned to the C-N stretching vibration of thiram, was selected to construct the intensity distributions. For thiram concentrations from  $1 \times 10^{-8}$  M to  $1 \times 10^{-6}$  M, the statistical SERS intensity distributions exhibited clear concentration-dependent features. As shown in Fig. 5(b),

the trained ResNet4 model achieved recognition accuracies higher than 95% for all tested concentrations, with an average accuracy of 99%.

To further examine the applicability of this method in a more complex biological matrix, we carried out concentration-level analysis of rosiglitazone maleate (ROS) in artificial serum. Before SERS measurements, serum samples were pretreated with methanol to precipitate proteins and reduce matrix interference. The corresponding SERS spectrum and peak assignments are shown in Fig. S14. Both characteristic Raman fingerprints of ROS and background signals from serum components were observed, reflecting the complexity of the biological matrix. The peak at  $739 \text{ cm}^{-1}$ , assigned to the benzene ring C-H deformation vibration of ROS, was selected to construct the intensity distributions. Despite the presence of serum-derived background signals, the statistical intensity distributions still showed concentration-dependent evolution as the ROS concentration increased from  $1 \times 10^{-7}$  M to  $1 \times 10^{-5}$  M. Based on these distribution patterns, the ResNet4 model achieved an average recognition accuracy of ~96%, as shown in Fig. 5(c). This result suggests that the proposed statistical SERS methodology can retain quantitative capability even in a complex biological matrix.

It should be noted that the present ResNet4 model

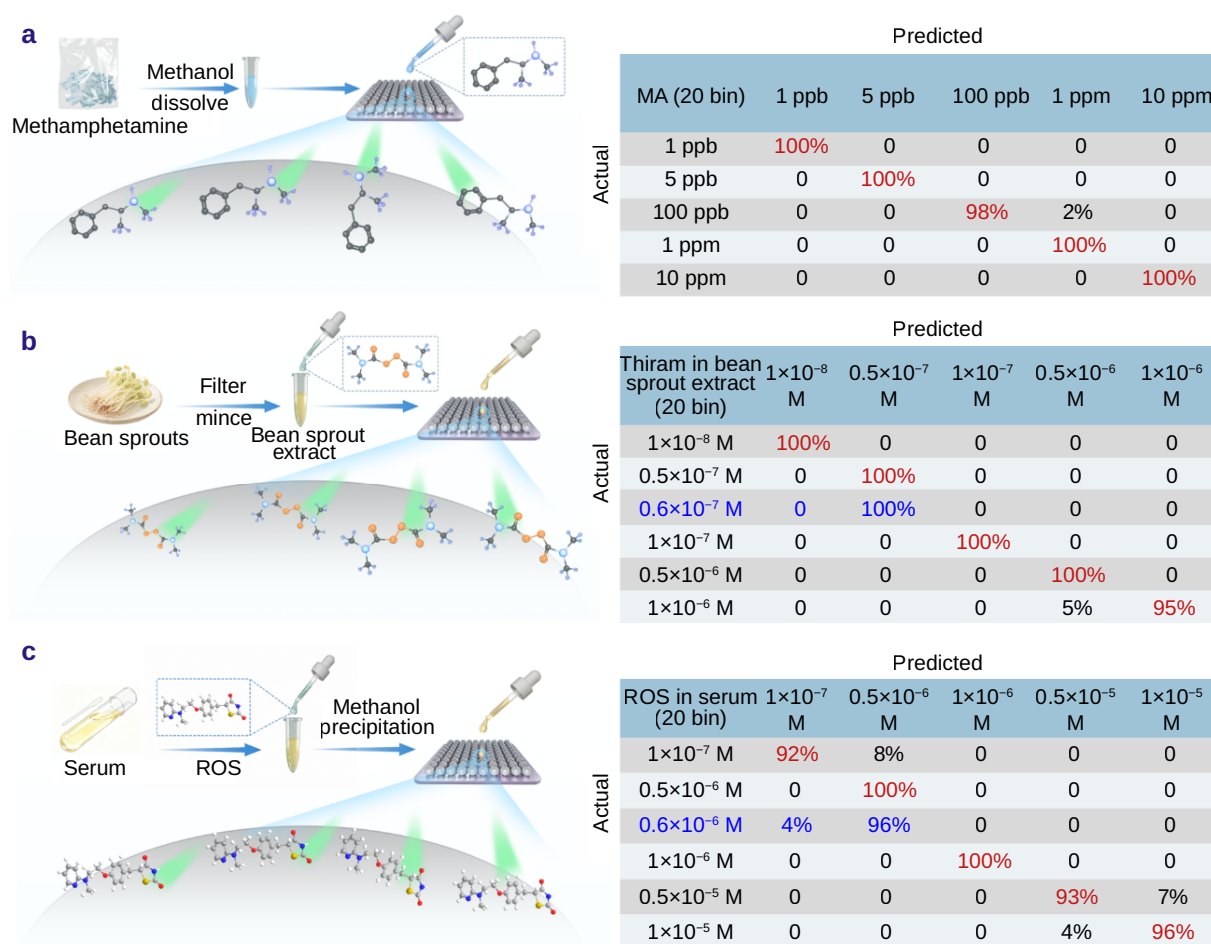


Fig. 5 | Applications of AI-driven statistical SERS quantitation. (a) Concentration-level identification accuracy of MA in methanol. (b) Concentration-level identification accuracy of thiram in bean sprout extract. (c) Concentration-level identification accuracy of ROS in methanol-treated serum.

performs concentration-level quantitation based on predefined concentration classes, rather than continuous regression prediction. To evaluate the response of the model to concentrations not included in the training set, several untrained intermediate concentrations were further tested. As shown in Fig. 5(b) and Fig. 5(c), the unknown samples at  $0.6 \times 10^{-7}$  M for thiram and  $0.6 \times 10^{-6}$  M for ROS were assigned mainly to their nearest trained concentration levels. This result indicates that the proposed statistical SERS strategy can estimate the concentration range of unknown samples within a predefined concentration grid. In principle, finer quantitative resolution can be achieved by introducing more concentration levels and sufficient statistical sampling data, as discussed in Table 2.

In addition, for all quantitative analysis, the training and testing datasets were derived from independent batches of SERS mapping measurements, further supporting the robustness of this statistical quantitation strategy against measurement-to-measurement and batch-to-batch variations. Taken together, the successful analysis of MA in

methanol, thiram in bean sprout extract, and ROS in methanol-treated serum demonstrate the generality of concentration-encoded SERS intensity distributions for reliable concentration-level quantitative analysis in different application scenarios.

### 3 Conclusions

In summary, we developed a statistical SERS quantitation strategy by combining concentration-encoded intensity distributions with deep-learning-assisted pattern identification. We demonstrated that SERS intensity distributions exhibit a clear concentration-dependent evolution, changing from asymmetric long-tailed profiles at low analyte concentrations to nearly symmetric Gaussian-like profiles at high concentrations. This evolution originates from stochastic molecular adsorption at plasmonic “hot spots” and can be reproduced by Poisson-statistics-based Monte Carlo simulations. By using the full intensity distributions as inputs, the ResNet4 model effectively identified

concentration-specific distribution patterns, enabling accurate concentration-level quantitation of R6G with an accuracy up to 100%. The applicability of this strategy was further verified by the analysis of methamphetamine in methanol, thiram in bean sprout extract, and rosiglitazone maleate in serum, demonstrating its potential for concentration-level quantitation in different analytical scenarios.

More importantly, this work extends statistical SERS quantitation beyond single-molecule digital counting. In digital SERS counting, analog SERS signals are typically converted into binary events, and the concentration is quantified from event probability based on Poisson statistics. In contrast, the present method preserves the full analog SERS intensity distribution as a concentration-encoded statistical fingerprint. This intensity-distribution metric captures richer statistical information, including molecular-number fluctuations, “hot spot” dependent enhancement variations, distribution skewness, and the overall evolution of the distribution profile. In this way, intrinsic SERS intensity fluctuations are transformed from a limitation of conventional intensity-based quantitation into useful statistical information. This work therefore proposes intensity distribution as a new statistical metric for SERS quantitation, deepens the understanding of stochastic nature of SERS spectroscopy, and provides a new route toward reliable SERS quantitative analysis beyond strict single-molecule conditions<sup>52–56</sup>.

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## Author contributions

This endeavor was accomplished through the collective efforts of all authors: Zhipeng Li and Hongxing Xu designed and supervised the research. Meng Zhang, Yue Wei and Longkun Yang performed the experiments. Hui Ding, Luyuan Xie and Shiyu Zhang carried out the deep learning model training/testing. Zhipeng Li, Chaowei Sun, Di Zheng, Ferruccio Pisanello and Longkun Yang analyzed the data. Longkun Yang, Zhipeng Li and Meng Zhang wrote the paper. All authors reviewed the paper.

## Competing interests

The authors declare no competing financial interests.

## Supplementary information

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